

PROCEEDINGS Open Access

A hybrid method for the exact planted (*l, d*) motif finding problem and its parallelization

Mostafa M Abbas^{1*}, Mohamed Abouelhoda^{2,3*}, Hazem M Bahig⁴

From Asia Pacific Bioinformatics Network (APBioNet) Eleventh International Conference on Bioinformatics (InCoB2012)

Bangkok, Thailand. 3-5 October 2012

Abstract

Background: Given a set of DNA sequences s_1 , ..., s_t , the (l, d) motif problem is to find an l-length motif sequence M, not necessary existing in any of the input sequences, such that for each sequence s_i , $1 \le i \le t$, there is at least one subsequence differing with at most d mismatches from M. Many exact algorithms have been developed to solve the motif finding problem in the last three decades. However, the problem is still challenging and its solution is limited to small values of l and d.

Results: In this paper we present a new efficient method to improve the performance of the exact algorithms for the motif finding problem. Our method is composed of two main steps: First, we process $q \le t$ sequences to find candidate motifs. Second, the candidate motifs are searched in the remaining sequences. For both steps, we use the best available algorithms. Our method is a hybrid one, because it integrates currently existing algorithms to achieve the best running time. In this paper, we show how the optimal value of q is determined to achieve the best running time. Our experimental results show that there is about 24% speed-up achieved by our method compared to the best existing algorithm. Furthermore, we also present a parallel version of our method running on shared memory architecture. Our experiments show that the performance of our algorithm scales linearly with the number of processors. Using the parallel version, we were able to solve the (21, 8) challenging instance using 8 processors in 20.42 hours instead of 6.68 days of the serial version.

Conclusions: Our method speeds up the solution of the exact motif problem. Our method is generic, because it can accommodate any new faster algorithm based on traditional methods. We expect that our method will help to discover longer motifs. The software we developed is available for free for academic research at http://www.nubios.nileu.edu.eg/tools/hymotif.

Background

DNA motifs are short sequences in the genome that play important functional roles in gene regulation. Due to their short length, it is difficult to identify these regions using features intrinsic in their composition. Assuming that the motifs are conserved in closely related species due to the importance of their function, it is possible to discover them by comparing the respective DNA

sequences to identify the sub-sequences that are very similar to each other.

There are two common combinatorial formulations that identify the motifs: The first is the consensus motif problem which made its first appearance in 1984 [1], while the second is the planted (l, d)-motif problem that was presented in 2000 [2]. It is worth noting that the latter formulation is a special case of the former. The exact definitions are as follows:

Given a set of t sequences s_i where $1 \le i \le t$ defined over an alphabet Σ . The consensus motif problem is to find an l-length motif sequence M such that in each sequence s_i , $1 \le i \le t$, there is at least one subsequence p_i

Full list of author information is available at the end of the article



^{*} Correspondence: mabbas@su.edu.eg; mabouelhoda@yahoo.com

¹Department of Basic Sciences, Faculty of Engineering, Sinai University, El-Arish, Edvot

²Systems and Biomedical Engineering Department, Faculty of Engineering, Cairo University, Giza, Egypt

differing with at most d mismatches from M; *i.e.*, $d_H(p_i, M) \le d$, where d_H is the hamming distance between p_i and M.

The planted (l, d) motif problem is a special case of the consensus problem in which we restrict that p_i occurs only once in s_i .

Due to its combinatorial nature, the consensus motif problem and its variant defined above is extremely challenging. Over a benchmark data of 20 sequences, each of length 600 characters, large instances of (15, 5), (17, 6), (19, 7) and (21, 8) have been addressed and many algorithms have been developed to solve them one after another. These algorithms can be classified into two major categories: approximation algorithms [2-12] and exact algorithms [13-30]. Approximation algorithms are based on probabilistic local search techniques, such as Gibbs Sampling, Expectation Maximization, etc. Although these algorithms may solve the challenging instances in practice, there is no guarantee that the motif can be found even when l is short.

Exact algorithms are based on exhaustive search techniques. The brute force algorithm proceeds by testing all possible motifs of length l using pattern matching, leading to $O(l \ n \ t \ 4^l)$ time complexity. This algorithm, however, is not suitable for discovering long motifs in practice, and many algorithms have been developed to provide faster solutions. Examples of these algorithms are CENSUS [23], PMS1 [26], PMSP [27], PMSprune [29], PMS5 [30], SMILE [19], RISO [24], RISOTTO [28], and Voting [25]. In the following we briefly review the most efficient ones and the ones related to our work.

The algorithms SMILE [19], RISO [24], and RISOTTO [28] are based on the use of suffix tree. The time complexity of these algorithms is the same and it is $O(t^2Nv(l, d))$,

where
$$v(l, d) = \sum_{i=0}^{d} C_i^l 3^i$$
 is the size of the *d*-mismatch

neighbourhood of motifs of length
$$l$$
 and $N = \sum_{i=1}^{l} n_i$, n_i is

the length of sequence i from input sequences. RISOTTO improved the time complexity of SMILE and RISO in the average case and solved some challenging instances such as (15, 5) and (17, 6).

PMSP [27] is based on exploring the neighbourhood of the l-mer of the first sequence and checking whether the elements of such neighbourhoods are (l, d) motifs. The time complexity is $O(\frac{1}{w}tn^2v(l,d))$. It is able to solve some challenging instances such as (15, 5) and (17, 6). PMSprune [29,31] is an improved version of the PMSP algorithm, based on the branch and bound strategy. Although it has the same worst-case time complexity as

PMSP algorithm, it is more efficient in practice and it could tackle the (17, 6) and (19, 7) instances for the first time. PMS5 [30] is based on computing the common *d*-neighbourhood of three *l*-mers using integer programming formulation. It combines this novel idea with the algorithms PMS1 and PMSPrune. PMS5 can tackle the large challenging instances (19, 7), (21, 8) and (23, 9). The only drawback of PMS5, it requires larger amount of internal memory to finish computation.

Our contribution

In a previous work [32,33], we have introduced an idea composed of two stages to speed up the exact algorithms: In the first stage, we generate a set of candidate motifs by applying one of the exact algorithms based on the neighbourhood method (like Voting [25] or PMSP [27] algorithms) using $q \le t$ sequences. In the second stage, for each candidate motif we check if it is a valid motif or not using pattern matching on the reminder (t - q) sequences. This dramatically reduces the search space and leads to significant speed up. The bottleneck in this approach, however, was the determination of the q value that yields the fastest running time. That is, the user has to guess the value of q, which might lead to non-optimal running time and even no speed up compared to the traditional methods. Also, the authors in [34] have used the same idea on PMS1, RISOTTO, and PMSprune algorithms.

In this paper, we present a theoretical method which can be used to determine the appropriate value of q. Then we apply this strategy on PMSprune algorithm and solve some big challenging instances such as (21, 8). Furthermore, we propose a parallel version of our algorithm to present a practical solution to the challenging instances of the motif problem. Our parallel version further speeds up the solution of the (21, 8) instance.

Definitions and related work

In this section, we introduce some notations and definitions that will help us to describe our algorithm and related work in a concise manner.

Definition 1 adapted from [29]: For any string x, with |x| = l, let $B_d(x) = \{y: |y| = l, d_H(y, x) \le d\}$, where d_H denotes the Hamming distance and $B_d(x)$ denotes the set of neighbourhoods of x. We also write v(l, d) to refer to $|B_d(x)|$.

Definition 2 adapted from [29]: Let s denote a string of length n and let x denote another string of length l, l < n. We define the minimum distance between s and x as $\bar{d}_H(x,s) = \min_{x' < q \mid s} d_H(x,x')$, where $x <_l s$ denotes that x is a substring of s with length l.

Definition 3 adapted from [29]: Given an l-length string x and a set of strings $S = \{s_1, ..., s_t\}$ with $|s_t| = n$

for i = 1, ..., t and l < n, we define the distance between S and x as $\bar{d}_H(x, S) = \max_{i=1}^t {\{\bar{d}_H(x, s_i)\}} = \max_{i=1}^t {\{\bar{d}_H(x, r)\}\}}$

Definition 4 adapted from [29]:A string x is an (l, d) motif for a set of sequences $S = \{s_1, ..., s_t\}$, if:

1) $\bar{d}_H(x,S) \leq d$.

2)
$$\exists y \triangleleft_l s_1 : x \in B_d(y) \land \bar{d}_H(x, \{s_1, ..., s_t\}) \leq d$$
.

Proposition 1 adapted from [10]: Let u and v be two random strings of length l over an alphabet of 4 characters with equal probability of occurrence. The probabil-

ity
$$p_d$$
 that $d_H(u, v) \le d$ is $p_d = \sum_{i=0}^d \binom{l}{i} (3/4)^i (1/4)^{l-i}$,

and the probability that $\bar{d}_H(x,S) \ge d$ is $(1-(1-p_d)^{n-l+1})^t$. The expected number of l-length motifs that occur at least once in each of the t sequences with up to d substitutions is $E(l, d, t, n) = 4^l(1-(1-p_d)^{n-l+1})^t$.

PMSprune Algorithm

Because the first stage of our method will depend on the PMSprune algorithm. We will review the basic steps of it in the notions presented above.

The main strategy of PMSprune is to generate $B_d(y)$, for every l-mer y in s_1 , using a branch and bound technique. An element $x \in B_d(y)$ is a motif only if $\overline{d}_H(x,S) \leq d$. The step of verifying that $\overline{d}_H(x,S) \leq d$ is achieved by scanning all substrings of S. For fixed values of t, n, and l, the expected time complexity of PMSprune is equal to

$$T_{PMSprune} = O\left(t(n-l+1)^2 \left(l + p_{2d} \sum_{i=1}^{2d-d'+1} {l \choose i} 3^i\right)\right)$$
 (1)

where p_{2d} is the probability that the hamming distance between two strings is at most 2d, and it is defined in Proposition 1. For fixed values of t, n, and l, value d' was estimated such that the probability of $\bar{d}_H(x,S) \geq d'$ is close to 1. (The probability of $\bar{d}_H(x,S) \geq d'$ is given in

Proposition 1 and it is
$$\left(1-\left(1-p_d'\right)^{n-l+1}\right)^t$$
).

Implementation

Our proposed strategy

Our new strategy, referred to as *hybrid exact pattern motif search* (HEP), is composed of three steps: first, we determine the value q, corresponding to the size of a subset of input sequences, as explained below. Second, we apply an exact exhaustive algorithm \pounds (like, PMSprune) on the set of q sequences to find the set of q-neighbourhood q-neighbourhood). We call this set the candidate motif set. Finally, we apply a pattern search algorithm over the remaining sequences to verify each

motif. Note that our algorithm is generic in the sense that it takes the program \pounds also as input in addition to the input sequences and user parameters. A pseudo code for this strategy using the exact algorithm \pounds is as follows:

Algorithm 2: HEP $(£, s_1,..., s_b, n, l, d)$ Begin

- 1) Determine the number of sequences q using the method given below.
- 2) Implement the exact algorithm \pounds on q input sequences. Let C be the set of candidate motifs found in the q sequences.
- 3) For each pattern ν in C, check if ν is a valid motif or not in the reminder (t q) input sequences using pattern matching Algorithm.

End.

Theorem 1: Algorithm 2 correctly finds all (l, d) motifs in a given t input sequences.

Proof: Step 2 of the algorithm is exhaustive and finds the whole set of d-neighborhood for the q sequences. Therefore, and by definition of the (l,d) motif problem, any (l,d) motif belongs to this set, even if q=1. In Step 3, each candidate motif is verified by comparison to each substring in the remaining sequences. This step is conducted by an approximate pattern matching algorithm for each l-length substring in the candidate motif set and each l-length substring in the remaining sequences such that the hamming distance between these two substrings is $\leq d$. This guarantees that no motif is missing.

Theorem 2: The running time of the HEP is equal to

$$T_{HEP} = T_{(q)} + l(t - q)(n - l + 1)E(l, d, q, n)$$
 (2)

where $T_{\mathcal{L}(q)}$ is the running time of step 2 involving the use of an exact algorithm \mathcal{L} on the q input sequences and l(t-q) (n-l+1) E(l,d,q,n) is the running time of step 3 such that E(l,d,q,n) is the number of elements in the set C, which is estimated to be $4l(1-(1-p_d)^{n-l+1})^q$. Note that the complexity of step 1 takes constant time, as we will explain below. Note that the running time of the brute force algorithm is acquired if q=0 in equation 2. The running time of the exact algorithm \mathcal{L} is acquired if q=t in equation 2.

Determination of the best q

The range of the number of sequences q, enhancing the performance of the exact motif finding problem is calculated by solving the following inequality for the parameter q:

$$T_{HEP} \le T$$
 (3)

Definition 5: We define *mns* as the minimum number of sequences q that yields better running time; i.e., the first value of q that verifies the inequality. We also define *ons* as the optimal number of sequences q that yields the best running time; i.e., the value of q such that T_{HEP} is minimum over $1 \le q \le t$.

Implementing HEP based on PMSprune

We decided to use PMSprune for implementing the first step in our method, because of its superiority compared to other algorithms as discussed in [31]. However, we stress that our approach is generic and can be used with any better algorithm that appears in future. In the following, we will refer to our method based on PMSprune as $HEP_PMSprune$. If q = mns we will denote it with $HEP_PMSprune(mns)$, and if q = ons we will denote it with $HEP_PMSprune(ons)$.

Determining mns for PMSprune

Replacing $T_{\mathcal{L}(q)}$ by the time of PMSprune on q sequences, Equations (1) and (2) can be rewritten as follows:

$$T_{PMSprune} = q(n-l+1)^{2} (l+p_{2d} \sum_{i=1}^{2d-d'+1} {l \choose i} 3^{i}) + (t-q)(n-l+1)^{2}$$

$$(l+p_{2d} \sum_{i=1}^{2d-d'+1} {l \choose i} 3^{i})$$

$$T_{HEP_PMSprune} = q(n-l+1)^2 (l+p_{2d} \sum_{i=1}^{2d-d'+1} \binom{l}{i} 3^i) + l(t-q)$$

$$(n-l+1)E(l,d,a,n)$$

Replacing T_{HEP} with $T_{HEP_PMSprune}$ and T_{\pounds} with $T_{PMSprune}$ in the inequality (3) results in the following variation:

$$l(t-q)(n-l+1)E(l,d,q,n) < (t-q)(n-l+1)^2(l+p_{2d}\sum_{i=1}^{2d-d'+1}\binom{l}{i}3^i)$$

Substituting the value of E(l, d, q, n) with the value given in Proposition 1 in the left hand side yields

$$4^{l} \left(1-(1-p_{d})^{n-l+1}\right)^{q} < \frac{(n-l+1)(l+p_{2d}\sum\limits_{i=1}^{2d-d'+1}\binom{l}{i}3^{i})}{l}$$

Dividing both sides by 4^l and taking the logarithm,

$$q > \frac{\log\left\{(n-l+1)(l+p_{2d}\sum_{i=1}^{2d-d'+1}\binom{l}{i}3^{i})\right\} - \log(l4^{l})}{\log(1-(1-p_{d})^{n-l+1})}$$
(4)

The inequality (4) provides the range of the values of q that makes the running time of HEP using PMSprune

less than the running time of the original PMSprune over the all set of sequences. The minimum value of q in the range of the inequality is called mns and it is equal to:

$$\left\lceil \frac{\log \left\{ (n-l+1)(l+p_{2d} \sum_{i=1}^{2d-d'+1} \binom{l}{i} 3^i) \right\} - \log(l4^l)}{\log(1-(1-p_d)^{n-l+1})} \right\rceil + 1$$

Determining ons for PMSprune

For fixed values of t, n, l and d, ons can be calculated for PMSprune by selecting the value of q that minimizes the total number of operations $T_{HEP_PMSprune}$ for $1 \le q \le t$. The following algorithm computes the value of ons for each instance (l, d).

Algorithm 3: Find ons

Begin

1)
$$q = ons = 1$$

2) $E(l, d, q, n) = 4^{l} (1 - (1 - (\sum_{i=0}^{d} {l \choose i} (3/4)^{i} (1/4)^{l-i}))^{n-l+1})^{q}$

3)
$$T_{\min} = q(n-l+1)^2(l+p_{2d}\sum_{i=1}^{2d-d'+1}\binom{l}{i}3^i) + l(t-q)(n-l+1)E(l,d,q,n)$$

4) for q = mns to t do

$$E(l,d,q,n) = 4^{l} \left(1 - \left(1 - \left(\sum_{i=0}^{d} \binom{l}{i} (3/4)^{i} (1/4)^{l-i}\right)\right)^{n-l+1}\right)^{q}$$

$$T = q(n-l+1)^2(l+p_{2d}\sum_{i=1}^{2d-d+1}\binom{l}{i}3^i) + l(t-q)(n-l+1)E(l,d,q,n)$$

if $T < T_{\min}$ then

 $T_{\min} = T$

ons = q

5) return ons

End

The above algorithm computes q in O(t) time. In practice, the time for computing q takes negligible time with respect to the rest of motif finding steps; it took maximum one second for all experiments included in this paper with simulated and real datasets. To save some time, our implementation includes a look-up table containing pre-computed values of q for different values of q, and q, where q is an and selected values of q with q is a solution of q in q i

Parallel version of HEP_PMSprune(ons)

We propose a parallel version for HEP_PMSprune(*ons*) called PHEP_PMSprune(*ons*). The two main steps of HEP_PMSprune(*ons*) can be parallelized as follows:

We parallelize the PMSprune algorithm by assigning a set of l-mers from s_1 to each processor for establishing the set of neighboring motifs. The resulting sets are stored in candidate motif lists C_i , $i \in \{1, 2, ..., p\}$, where p is the number of processors. After each processor finishes computation, the C_i lists are merged together in a larger set C, such that each motif is represented once in this list; i.e., all repetitions are removed. Creating the C list is done in linear time with respect to the number of candidate motifs and it is achieved as follows:

We incrementally construct the partial list C_i that contains the L_i lists, $1 \le j \le p$, by appending the list L_i at the end of the list C_{i-1} such that all elements in L_i existing in C_{j-1} are discarded. This continues until j = p; i.e., C_p is C. Discarding a repeated element is done efficiently as follows: For small values of l, we create a look-up table with size Σ^l , where Σ is the alphabet size. Each possible *l*-length string can be mapped to a number in the range between zero and Σ^l in O(l) time. The i^{th} entry in this table contains one if a string in C_{i-1} is mapped to i. Otherwise, it contains zero. The strings in C_i are queried against this look-up table to discard repetitions and set entries they are mapped to with value one. For longer values of l, we use the Aho-Corasick automaton to index all l-length motifs in C_{j-1} , and check if a strings in C_i exists in the automaton or not and add the new strings of C_i to the automaton. For these string matching algorithms, we refer the reader to [35].

In the second step, we validate each candidate motif independently in parallel over the available processors. The running time of this algorithm is $O(T_s/p + |C|)$, where T_s is the sequential running time and |C| is the size of set C.

The first step in the parallel algorithm does not lead to loss of any motifs. This is because the set C includes the d-neighborhood set of the q-sequences. The reason is that we run PMSprune in parallel against the strings $(x, s_2, s_3, ..., s_q)$, where x is a substring of s_1 . That is, each substring is not processed. The second step in the parallel algorithm is also correct, because the elements in C are independent of each other and checking the validity of each candidate motif can be safely run in parallel. Our experimental results confirm the correctness of our parallelization procedure.

Results and discussion

Experiments on simulated datasets

We used the simulated data sets that are used in many articles [25-30,32-34] with t = 20 sequences and n = 600characters, where the alphabet size is 4. Each (*l*, *d*) input instance dataset is generated as follows: We generate random strings with length (n-l) each, where the characters appear randomly with equal probability. Then we generate randomly an l-length string M and plant a copy of it in each sequence at random position after mutating it with at most d random mutations. We tested the algorithms for varying n, l, and d values and for the following challenging instances: (11, 3), (13, 4), (15, 5), (17, 6), (19, 7), and (21, 8).

Experiments overview

Our experiments address three major issues: The first is the performance of our method compared to the use of PMSprune only. The second, we show that our method for selecting q, already achieves the best running time. The third is the performance of the parallel version and its scalability. The algorithms are implemented on a 2 Quad-core processors (2.5 GHz each) machine. The programs are coded in C language. In the parallel version, we use openMP directives for parallelizing the code.

Performance of HEP on PMSprune

Tables 1 and 2 show the performance of the algorithms HEP_PMSprune(mns) and HEP_PMSprune(ons) with respect to PMSprune algorithm respectively. The last column in Tables 1 and 2 displays the improvement in

PMSprune which equals to
$$\frac{T_{PMSprune} - T_{HEP_PMSprune(mns)}}{T_{PMSprune}}$$

column in Tables 1 and 2 displays the improvement in PMSprune which equals to
$$\frac{T_{PMSprune} - T_{HEP_PMSprune(mns)}}{T_{PMSprune}}$$
 and
$$\frac{T_{PMSprune} - T_{HEP_PMSprune(ons)}}{T_{PMSprune}}$$
 respectively. We used the notations $\binom{n'}{n}$ $\binom{n'}{n}$ $\binom{n'}{n}$ and $\binom{n'}{n}$ in computing the time

the notations 's', 'm', 'h', and 'dy' in computing the time for seconds, minutes, hours, and days, respectively. The results confirm that, the algorithms HEP_PMSprune (mns) and HEP_PMSprune(ons) significantly reduced the running time compared to the standard PMSprune algorithm in all challenging instances.

Evaluating the choice of q

In this section, we experimentally evaluate our algorithm for determining the best q that minimizes the running time of the $HEP_PMSprune(q)$ algorithm. To achieve this, we will follow the following steps:

1. We run HEP_PMSprune(q), $mns \le q \le t$ for the problem instances (11, 3), (13, 4), (15, 5), (17, 6), (19, 7),

Table 1 Time Comparison of PMSPrune and HEP_PMSprune(mns) with the Challenging Instances

1	d	T _{PMSprune}	mns	$T_{HEP_PMSprune}(_{mns})$	Improvement
11	3	1.92 s	9	1.4 s	27.1 %
13	4	33.95 s	7	26.05 s	23.27 %
15	5	7.7 m	6	6.4 m	16.8 %
17	6	1.55 h	7	1.26 h	18.5 %
19	7	18.62 h	6	14.93 h	19.8 %
21	8	8.59 dy	6	6.68 dy	22.23 %

Table 2 Time Comparison of PMSPrune and HEP_PMSprune(ons) with the Challenging Instances

I	D	T _{PMSprune}	ons	$T_{HEP_PMSprune}(ons)$	Improvement
11	3	1.92 s	10	1.34 s	30 %
13	4	33.95 s	9	24.55 s	27.69 %
15	5	7.7 m	7	6.02 m	21.8 %
17	6	1.55 h	8	1.26 h	18.65 %
19	7	18.62 h	7	14.39 h	22.74 %
21	8	8.59 dy	6	6.68 dy	22.23 %

and (21, 8) and determine the value of q that minimizes the running time; we will refer to this value with ons_{exp} .

2. Compare the ons_{exp} against our ons computed theoretically.

Figure 1, which plots the running time against different q values, shows the results of applying these steps. We observe the value of *ons* is equal or very close to the value of $ons_{\rm exp}$.

We also conducted another experiment, where the problem instances were generated with different n and l and d. Table 3 shows the results for many of these instances, where the number of sequences t=20. We can observe that our algorithm finds the optimal q in all these instances. We also observe improvement of the running time with respect to the PMSprune algorithm in most of the cases. The cases with no improvement in the running time are attributed to the fact that the expected number of motifs is very low and the original algorithm runs already fast in these cases.

Note that it was not feasible to list the results for all possible values n, l, and d in Table 3. But in other instances with different values of n, l, and d, we found that ons and its time were consistent with $ons_{\rm exp}$ and its time published in this table.

Performance of PHEP_PMSprune(ons)algorithm

In Table 4, we show the results of applying the parallel version of our algorithm PHEP_PMSprune(ons) using different number of processors and for different problem instances. The running time of the difficult instance (21, 8) has been decreased from 6.68 days to about 20.42 hours using 8 processors. Figure 2 shows the scalability results for the algorithm where

speedup =
$$\frac{T_{HEP_{PMSprune(ons)}}}{T_{HEP_{PMSprune(ons)}}}$$
. From Table 4 and Figure 2 we

note that PHEP_PMSprune(ons) reduce the time of HEP_PMSprune(ons) and the speedup achieved scales well with the increasing number of processors.

Experiments on real datasets

We used two collections of real datasets used in previous research papers [10,26,29,36]. The first collection

is a dataset including a number of the upstream regions of yeast genes [37] affected by certain transcription factors. The transcription factors are from the SCPD [38] database and the paper [39]. The upstream DNA sequences were extracted using the Saccharomyces Genome Database [37]. The second collection includes the dataset of Blanchette [36] which includes the upstream DNA regions of many genes from different species. This dataset is available at http://bio.cs.washington.edu/supplements/FootPrinter and a copy of it is available with our software tool for testing.

Tables 5 and 6 show the motifs found by our method compared to the published ones for both collections. In each table, we give a reference to the published motif. Our program could detect all published motifs. It is also interesting to note that our program could detect extra novel motifs in the case of the Interleukin-3 problem instance in Table 6. These motifs look interesting, because they are 20 bp long with hamming distance zero; an observation that calls for further biological investigation.

Tables 5 and 6 also include the running times (in seconds) of running our method for the listed problem instances and the improvement in time compared to the PMSprune method. The running time for one problem instance is the time needed to run our program in the $(l,\ d)$ parameters range from $(6,\ 0)$ until $(21,\ 3)$, i.e., there are 64 invocations of our program. The results show that our program is superior to the PMSprune for large instances.

Conclusions

In this paper, we introduced an efficient method that can enhance the performance of exact algorithms for the motif finding problem. Our method depends on dividing the sequence space into two sets. Over the first set, we generate a set of candidate motifs. Then, we use the remaining set of sequences to verify if each candidate motif is a real one. The experimental results show that our method is superior to the best methods available so far and could tackle large problems like (21, 8). Finally, we introduced a scalable and efficient parallel version for the proposed method. Our tool is available for free for academic research at http://www.nubios.nileu.edu.eg/tools/hymotif.

Availability and requirements

Project name: hymotif.

Project home page: http://www.nubios.nileu.edu.eg/tools/hymotif

Operating system(s): Linux. Programming language: C.

Other requirements: C/C++ libraries.

License: GPL.

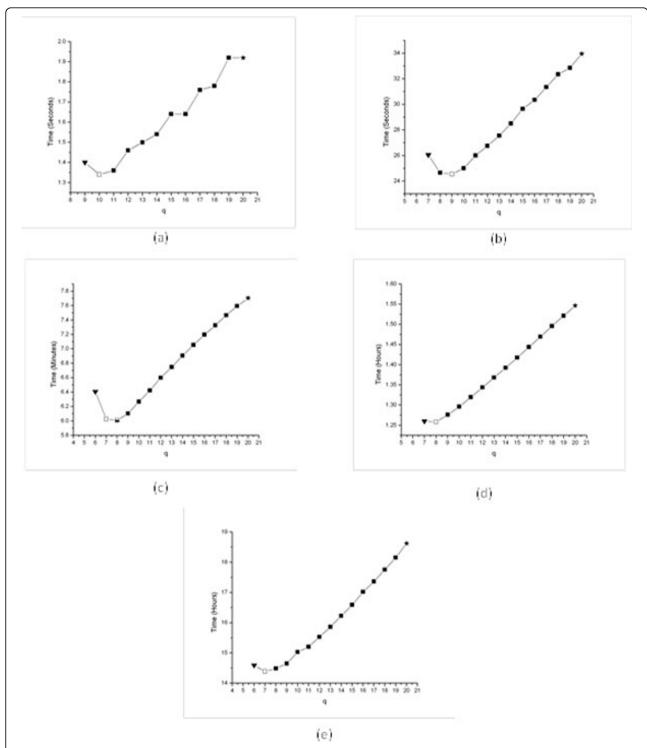


Figure 1 Performance of our method for different challenging instances. Behavior of HEP_PMSprune(q) for different (l, d) instances such that $q \in \{mns, ..., t\}$. (a): (11, 3), (b): (13, 4), (c): (15, 5), (d): (17, 6), (e): (19, 7). We used the following remarks in the figures: 1) Black-triangle-down to indicate the running time of HEP_PMSprune(mns). 2) Black-star to indicate the running time of PMSprune or HEP_PMSprune(mns): i.e., using theoretically estimated q.

Table 3 The performance of the HEP_PMSprune(ons) for different values of n and l

n	d	I	ons	T_ons	ons_exp	T_onsexp	T_pms
300	3	11	9	0.0001	3-20	0.0001	0.0001
600	3	11	10	1.34	10	1.34	1.92
900	3	11	14	4	11-16	4	5
1200	3	11	17	7	17	7	8
1500	3	11	20	16	20	16	16
300	3	12	6	0.05	4-20	0.05	0.05
600	3	12	8	0.83	4-20	0.83	0.83
900	3	12	8	1.5	6-20	1.5	1.5
1200	3	12	9	3	6-15	3	4
1500	3	12	10	5	8-12	5	7
300	4	13	7	3	5-20	3	3
600	4	13	9	24.55	9	24.55	33.95
900	4	13	11	81	11	81	109
1200	4	13	14	190	14	190	217
1500	4	13	17	353	17-19	356	360
300	4	14	6	1	4-20	1	1
600	4	14	7	6.5	7-18	6.5	7
900	4	14	8	21.5	8-9	21.5	24
1200	4	14	8	54	8	54	67
1500	4	14	9	107	9	107	146
300	4	15	5	0.25	4–20	0.25	0.25
600	4	15	5	1.25	4-20	1.25	1.25
900	4	15	6	5	5-20	5	5
1200	4	15	6	12	8	10	13
1500	4	15	7	16.5	7-13	16.5	20
300	4	16+	5	0.002	3-20	0.002	0.002
600	4	16+	5	0.25	4-20	0.25	0.25
900	4	16+	5	1	4-20	1	1
1200	4	16+	6	2.34	5-20	2.34	2.34
1500	4	16+	6-8	4.89	5-20	4.89	4.89
300	5	15	7	38	6-10	38	46
600	5	15	8	361.2	8	360	462
900	5	15	9	1250	9	1250	1847
1200	5	15	11	2976	11	2976	4060
1500	5	15	13	5829	13	5829	6969
300	5	17	5	2	5-20	2	2
600	5	17	6	27	13-20	19	19
900	5	17	5	103	7-20	92	92
1200	5	17	6	231	6-8	224	264
1500	5	17	6	439	6-8	439	552
300	5	18+	5	1	5-20	1	1
600	5	18+	6	5	6-20	4	4
900	5	18+	6-7	14	6-20	14	14
1200	5	18+	6-7	33	6-20	33	33
1500	5	18+	6-8	74	6-20	74	74

The first column includes the sequence length *n*, the second includes the hamming distance *d*, and the third includes the motif length *l*. The entries *l*+, means greater than *l* leads to no improvement. 'ons' stands for the theoretically computed *q*, while "ons_exp" stands for the experimentally found one. We report range of ons_exp that yielded best time. There also range of ons for *l*+. "T_ons" and "T_ons_{exp}" stand for the times (in seconds) with ons and ons_exp, respectively. "T_pms" stands for the time with the original PMSprune algorithm only.

Table 4 Running time of PHEP_PMSprune(ons) using different number of processors p for some challenging instances

1	d	Time							
			P = 2	P=3	P =4	<i>P</i> = 5	<i>P</i> = 6	P = 7	<i>P</i> = 8
13	4	24.86 s	12.4 s	8.35 s	6.1 s	4.95 s	4.35 s	3.6 s	3.2 s
15	5	6.34 m	3.19 m	2.13 m	1.61 m	1.28 m	1.07 m	55.2 s	48.5 s
17	6	1.28 h	38.28 m	25.58 m	19.16 m	15.34 m	12.81 m	10.98 m	9.61 m
19	7	14.56 h	7.24 h	4.81 h	3.61 h	2.98 h	2.42 h	2.07 h	1.82 h
21	8	6.68 dy	3.33 dy	2.23 dy	1.67 dy	1.34 dy	1.12 dy	23.18 h	20.42 h

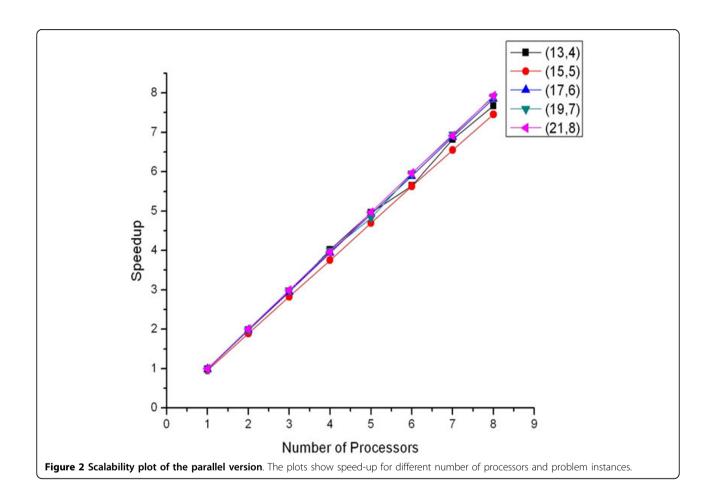


Table 5 Application of the PHEP_PMSprune(ons) on the real yeast dataset

Transcription Factor	Genes	Detected motif (s) & parameters	Published Motif (s) & reference(s)	Time
PHO4 (600 bp)	PHO5, PHO8, PHO81, PHO84,	CACGTG (6,0)	CACGT[G T] [38]	38 (5%)
HSE_HSTF (600 bp)	SSA1, HSP26, SSA4, HSC82, SIS1, CUP1-1	TTCAGTGAA (9,2)	TTCNNGAA [38] TTCNNNGAA [38]	37 (35%)
PDR (600 bp)	PDR3, SNQ2, PDR15, HXT9, HXT11, PDR5, YOR1	TCCGTGGA (8,1) TCCGCGGA (8,1)	TCCG[C T]GGA [38]	27(13%)
MCB (600 bp)	CDC2, CDC9, CDC6, CLN1, POL1, CDC21	ACGCGT (6,0)	[A T]CGCG[A T] [38]	31(20%)

Table 5 Application of the PHEP_PMSprune(ons) on the real yeast dataset (Continued)

ECB	SWI4, MCM5	TTTCCCATTAAGGAAA (16,3)	TTtCCcnntnaGGAAA [10,39]	41(49%)
(600 bp)	MCM7, CDC6			
	CLN3			

The first column includes the transcriptional factors (regulatory elements) and the length of upstream sequences. The second column includes the regulated genes. The first three factors and their related genes are available at the SCPD [38]. The ECB is the *early-cell-cycle-box* promoter region described in [39] and we extracted its related genes from the Yeast Genome Database [37]. The third column includes the motif detected by our tool and the respective parameters (*l*, *d*). The fourth column includes the published motifs and their references. The final column includes the running time in seconds needed to run our program in the parameter range from (6, 0) until (21, 3), i.e., there are 64 invocation of our program. The percentages in brackets refer to percentage improvements in rum time compared to PMSprune method.

Table 6 Application of the PHEP_PMSprune(ons) on the Blanchette real dataset

DNA region	Seq. no.	Detected motif	Published Motif	Time
Insulin family 5' promoter (500 bp)	8	CCTCAGCCCC (10, 1)	CCTCAGCCCC [10,40]	87(10%)
		AAGACTCTAA (10,2)	AAGACTCTAA [36,40]	
		GCCATCTGCC (10,1)	GCCATCTGCC [36,40]	
		CTATAAAG (8,0)	CTATAAAG [36, GB]	
	<u> </u>	GGGAAATG (8,1)	GGGAAATG [36,40]	
Metallothionein 5'UTR+Promoter (590 bp)	26	TTTGCACACGC (11,3)	TTTGCACACG [36,40]	7.87(1%)
		TGCACAC (7,1)	TGCACACGG [36,40]	
nterleukin-3 5'UTR+Promoter 490 bp	6	TTGAGTACT (9,2)	TTGAGTACT [36,40]	
		GATGAATAAT (10,1)	GATGAATAAT [36,40]	
		TCTTCAGAG, (9,2)	TCTTCAGAG [36,40]	
		AGGACCAG, (8,1)	AGGACCAG [36,40]	466(10%)
		AGGTTCCATGTCAGATAAAG, ATGGAGGTTCCATGTCAGAT, CTATGGAGGTTCCATGTCAG, GAGGTTCCATGTCAGATAAA, GGAGGTTCCATGTCAGATAAA, TATGGAGGTTCCATGTCAGATAA, TGGAGGTTCCATGTCAGATA, all these motifs found with (20,0)	Novel	
Growth-hormone 5 UTR+promoter (380 bp)	16	AACTTATCCAT (11,3)	ATTATCCAT [36,40]	3.43(0%)
		ATAAATGTAAA (11,3)	ATAAATGTA [36,40]	
		TATAAAAAG (9,2)	TATAAAAAG [36,40]	
c-fos 5 [°] UTR+promoter (800bp)	6	CCATATTAGGAC (12,3)	CCATATTAGGACATCT [10,41]	350(15%)
		GAGTTGGCTGC (11,3)	GAGTTGGCTG [36]	
		CACAGGATGT (10,2)	CACAGGATGT [36,40]	
		AGGACATCTGCT (12,3)	AGGACATCTG [36,40]	
c-myc 5'+promoter (100bp)	7	GTTTATTC (8,1)	GTTTATTC [36]	83.5(42%)
		CTTGCTGGG (9,2)	TTGCTGGG [36]	
		TGTTTACATC (10,2)	TGTTTACATC [36,40]	
		CCCTCCCC (8,1)	CCCTCCCC [36,40]	

Table 6 Application of the PHEP PMSprune(ons) on the Blanchette real dataset (Continued)

Histone H1 5'UTR+Promoter 650 bp	4	CAATCACCAC, (10,2)	CAATCACCAC, [36, GB]	47.6(9%)
		AAACAAAAGT (10,1)	AAACAAAAGT, [36, GB]	

The first column includes the gene family and the length of upstream sequences. The second column includes the number of sequences. The third column includes the motif detected by our tool and the respective parameters (*l*, *d*). The fourth column includes the published motifs and their references; "GB" stands for Genebank annotation. The final column includes the running time in seconds needed to run our program in the parameter range from (6,0) until (21,3), i.e., there are 64 invocation of our program. The percentages in brackets refer percentage improvements in rum time compared to PMSprune method.

Any restrictions to use by non-academics: No restrictions.

Acknowledgements

The authors are grateful to M.M. Mohie Eldin for useful discussion. The authors also thank Sanguthevar Rajasekaran for providing us with the source code of PMSprune and real datasets.

This article has been published as part of *BMC Bioinformatics* Volume 13 Supplement 17, 2012: Eleventh International Conference on Bioinformatics (InCoB2012): Bioinformatics. The full contents of the supplement are available online at http://www.biomedcentral.com/bmcbioinformatics/supplements/13/S17.

Author details

¹Department of Basic Sciences, Faculty of Engineering, Sinai University, El-Arish, Egypt. ²Systems and Biomedical Engineering Department, Faculty of Engineering, Cairo University, Giza, Egypt. ³Center for Informatics Sciences, Nile University, Giza, Egypt. ⁴Computer Science Division, Department of Mathematics, Faculty of Science, Ain ShamsUniversity, Cairo 11566, Egypt.

Authors' contributions

All authors contributed to theoretical and practical developments which form the basis of HEP method. All authors wrote and approved the manuscript.

Competing interests

The authors declare that they have no competing interests.

Published: 13 December 2012

References

- Waterman MS, Aratia R, Galas DJ: Pattern recognition in several sequences: consensus and alignment. Bulletin of Mathematical Biology 1984, 46(4):515-527.
- Pevzner P, Sze SH: Combinatorial approaches to finding subtle signals in DNA sequences. Proceedings of Eighth International Conference on Intelligent Systems for Molecular Biology 2000, 269-278.
- Lawrence C, Reilly A: An expectation maximization (EM) algorithm for the identification and characterization of common sites in unaligned biopolymer sequences. Proteins: Structure, Function and Genetics 1990, 7:41-51.
- Lawrence C, Altschul S, Boguski M, Liu J, Neuwald A, Wootton J: Detecting subtle sequence signals: A Gibbs Sampling strategy for multiple alignment. Science 1993, 262:208-214.
- Bailey T, Elkan C: Unsupervised learning of multiple motifs in biopolymers using Expectation Maximization. Machine Learning 1995, 21:51-80.
- Fraenkel Y, Mandel Y, Friedberg D, Margalit H: Identification of common motifs in unaligned DNA sequences: application to Escherichia coli Lrp regulon. Bioinformatics 1995, 11:379-387.
- Rigoutsos I, Floratos A: Combinatorial pattern discovery in biological sequences: the TEIRESIAS algorithm. Bioinformatics 1998, 14:55-67.
- Hertz GZ, Stormo GD: Identifying DNA and protein patterns with statistically significant alignments of multiple sequences. *Bioinformatics* 1999. 15:563-577.
- Gelfand M, Koonin E, Mironov A: Prediction of transcription regulatory sites in archaea by a comparative genomic approach. Nucleic Acid Res 2000, 28:695-705.

- Buhler J, Tompa M: Finding motifs using random projections. Journal of Computational Biology 2002, 9(2):225-242.
- 11. Price A, Ramabhadran S, Pevzner PA: Finding subtle motifs by branching from sample strings. *Bioinformatics* 2003, 19(2):ii149-ii155.
- Huang C, Lee W, Hsieh S: An improved heuristic algorithm for finding motif signals in DNA sequences. IEEE/ACM Transactions on Computational Biology and Bioinformatics 2011, 8(4):959-975.
- Galas DJ, Eggert M, Waterman MS: Rigorous pattern-recognition methods for DNA sequences: analysis of promoter sequences from Escherichia coli. Journal of Molecular Biology 1985, 186(1):117-128.
- Staden R: Methods for discovering novel motifs in nucleic acid sequences. Computer Applications in the Biosciences 1989, 5(4):293-298.
- Brazma A, Jonassen I, Vilo J, Ukkonen E: Predicting gene regulatory elements in silico on a genomic scale. Genome Research 1998, 15:1202-1215.
- Sagot MF: Spelling approximate repeated or common motifs using a suffix tree. In Latin'98: Theoretical informatics, LNCS Lucchesi CL, Moura AV 1998, 1380:111-127.
- Van Helden J, Andre B, Collado-Vides J: Extracting regulatory sites from the upstream region of yeast genes by computational analysis of oligonucleotide frequencies. *Journal of Molecular Biology* 1998, 281(5):827-842.
- Tompa M: An exact method for finding short motifs in sequences with application to the ribosome binding site problem. Proceedings of seventh International Conference on Intelligent Systems for Molecular Biology 1999, 262-771
- Marsan L, Sagot MF: Algorithms for extracting structured motifs using a suffix tree with an application to promoter and regulatory site consensus identification. *Journal of Computational Biology* 2000, 7(3-4):345-362.
- Sinha S, Tompa M: A statistical method for finding transcription factor binding sites. Proceedings of Eighth International Conference on Intelligent Systems for Molecular Biology 2000, 344-354.
- Blanchette M: Algorithms for phylogenetic footprinting. Proceedings of Fifth International Conference Computational Biology (RECOMB 2001) 2001.
- Eskin E, Pevzner PA: Finding composite regulatory patterns in DNA sequences. Bioinformatics 2002, 18(1):354-363.
- Evans PA, Smith A: Toward optimal motif enumeration. Proceedings of Eighth International Workshop Algorithms and Data Structures (WADS03) 2003. 47-58.
- Carvalho AM, Freitas AT, Oliveira AL, Sagot MF: A highly scalable algorithm for the extraction of CIS-Regulatory regions. Proceedings of Third Asia Pacific Bioinformatics Conference 2005, 273-282.
- Chin FYL, Leung HCM: Voting algorithms for discovering long motifs. Proceedings of Third Asia Pacific Bioinformatics Conference 2005, 261-271.
- Rajasekaran S, Balla S, Huang C-H: Exact algorithms for planted motif problems. Journal of Computational Biology 2005, 12(8):1117-1128.
- Davila J, Balla S, Rajasekaran S: Space and time efficient algorithms for planted motif search. Proceedings of Second International Workshop on Bioinformatics Research and Applications (LNCS 3992) 2006, 822-829.
- Pisanti N, Carvalho A, Marsan L, Sagot MF: RISOTTO: fast extraction of motifs with mismatches. Proceedings of Seventh Latin American Theoretical Informatics Symposium 2006, 757-768.
- Davila J, Balla S, Rajasekaran S: Fastand practical algorithms for planted (I, d) motif search. IEEE/ACM Transactions on Computational Biology and Bioinformatics 2007, 544-552.
- Dinh H, Rajasekaran S, Kundeti V: PMS5: an efficient exact algorithm for the (I, d)-motif finding problem. BMC Bioinformatics 2011, 12:410-420.
- Sharma D, Rajasekaran S, Dinh H: An experimental comparison of PMSprune and other algorithms for motif search. CoRR abs 2011, 1108.5217.

- Abbas MM, Bahig HM: Performance and analysis of modified voting algorithm for planted motif search. Proceedings of Seventh ACS/IEEE International Conference on Computer Systems and Applications 2009, 775-731
- Bahig HM, Abbas MM, Bhery A: Experimental study of modified voting algorithm for planted (I, d)-motif problem. Experimental Medicine and Biology, special issue Advanced in Computational Biology, Springer 2010, 65-73.
- Rajasekaran S, Dinh H: A speedup technique for (I, d)-motif finding algorithms. BMC Research Notes 2011, 4: 54-61.
- Gusfield D: Algorithms on strings, trees, and sequences: computer science and computational biology. Cambridge university Press, New York 1977
- Blanchette J, Tompa M: Discovery of regulatory elements by a computational method for phylogenetic footprinting. Genome Research 2002, 12(5):739-748.
- 37. Cherry JM, Adler C, Ball C, Chervitz SA, Dwight SS, Hester ET, Jia Y, Juvik G, Roe T, Schroeder M, Weng S, Botstein D: SGD: Saccharomyces Genome Database. Nucleic Acid Res 1998, 26:73-79 [http://www.yeastgenome.org/].
- Zhu J, Zhang M: SCPD: A Promoter Database of the Yeast Saccha-Romyces Cerevisiae. Bioinformatics 1999, 15(7-8):607-611[http://cgsigma.cshl.org/jian/].
- McInerny CJ, Partridge JF, Mikesell GE, Creemer DP, Breeden LL: A novel Mcm1-dependent element in the SWI4, CLN3, CDC6, and CDC47 promoters activates M/G1-specific transcription. Genes Dev 1997, 11(10):1277-1288.
- Wingender E, Dietze P, Karas H, Knuppel R: TRANSFAC: a database on transcription factors and their DNA binding sites. Nucleic Acids Research 1996, 24(1):238-241[http://transfac.gbf-braunschweig.de/TRANSFAC/].
- Natesan S, Gilman M: YY1 facilitates the association of serum response factor with the c-fos serum response element. Mol Cell Biol 1995, 15(11):5975-5982.

doi:10.1186/1471-2105-13-S17-S10

Cite this article as: Abbas *et al.*: A hybrid method for the exact planted (*l, d*) motif finding problem and its parallelization. *BMC Bioinformatics* 2012 13(Suppl 17):S10.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

