CORRECTION Open Access

# Correction: MGcount: a total RNA-seq quantification tool to address multi-mapping and multi-overlapping alignments ambiguity in non-coding transcripts

Andrea Hita<sup>1,2</sup>, Gilles Brocart<sup>1</sup>, Ana Fernandez<sup>1,2</sup>, Marc Rehmsmeier<sup>2</sup>, Anna Alemany<sup>3†</sup> and Sol Schvartzman<sup>1\*†</sup>

The original article can be found online at https://doi.org/10.1186/s12859-021-04544-3.

<sup>†</sup>Anna Alemany and Sol Schvartzman contributed equally to this work

\*Correspondence: sol.schvartzman@diagenode. com

<sup>1</sup> Epigenetics Unit, Diagenode s.a., Liège, Belgium Full list of author information is available at the end of the article

### Correction to: BMC Bioinformatics (2022) 23:39

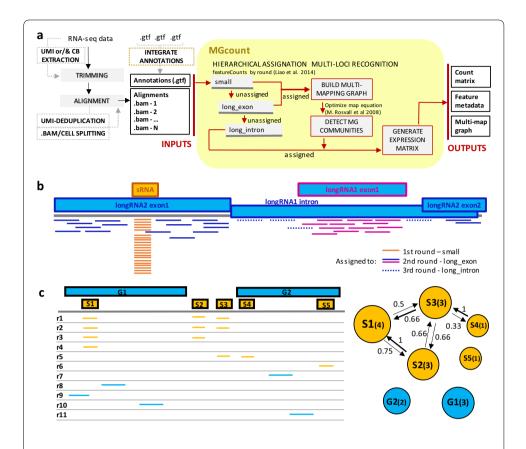
https://doi.org/10.1186/s12859-021-04544-3

Following the publication of the original article [1], the authors identified an error in Fig. 2 and caption 2c. The correct figure is given below, and the caption has been updated from "Reads ri (i = 1, 10)" to "Reads ri (i = 1, 11)."

The original article [1] has been corrected.



Hita et al. BMC Bioinformatics (2022) 23:209 Page 2 of 2



**Fig. 2** MGcount strategy. **a** MGcount takes a set of genomic alignments (BAM files) and a GTF RNA feature annotations file as inputs. The algorithm assigns reads hierarchically and then models multi-mapping assignments in a graph using the Rosvall's map equation [36, 37]. As output, MGcount provides an RNA expression count matrix (where feature communities are collapsed as new defined features), a feature metadata table and the graphs. **b** Illustration of how the hierarchical assignation can resolve multi-overlappers: reads that map to small-RNA and long-RNA features are assigned to small-RNA in the first round; reads that map to long-RNA introns and long-RNA exons are assigned to long-RNA exons in the second round; remaining reads are assigned in the last round. **c** Illustration of multi-mapping small-RNA and long-RNA exon graphs generation by MGcount. Reads ri (i = 1, 11) have been hierarchically assigned to  $S_1$ ,  $S_2$ ,  $S_3$ ,  $S_4$ ,  $S_5$  (small-RNA biotypes, yellow), and  $G_1$ ,  $G_2$  (long-RNA biotypes, blue). Each vertex in the directional multi-mapping graphs (right) corresponds to a feature and has a size proportional to the logarithm of the number of alignments. Edges connect vertices with common multi-mapping reads, with weights proportional to the number of common multi-mappers normalized by the total number of alignments of the source vertex. Hence, the weight of the edge connecting S1 with S2 becomes 3/4 (reads mapping both S1 and S2 divided by reads aligned to S1). (CB: Cell Barcode, UMI: Unique Molecular Identifier)

## **Author details**

<sup>1</sup>Epigenetics Unit, Diagenode s.a., Liège, Belgium. <sup>2</sup>Department of Biology, Humboldt-Universität Zu Berlin, Germany. <sup>3</sup>Department of Anatomy and Embryology, Leiden University Medical Centre, Leiden, The Netherlands.

### Published online: 01 June 2022

### Reference

 Hita, et al. MGcount: a total RNA-seq quantification tool to address multi-mapping and multi-overlapping alignments ambiguity in non-coding transcripts. BMC Bioinformatics. 2022;23:39. https://doi.org/10.1186/ s12859-021-04544-3.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.